

Introduction to RBM package

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1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 20

> which(myresult$permutation_p<=0.05)

[1] 5 22 67 72 188 249 318 395 403 420 504 617 671 721 736 756 824 832 850
[20] 960

> sum(myresult$bootstrap_p<=0.05)

[1] 16

> which(myresult$bootstrap_p<=0.05)

[1] 26 95 223 347 411 450 466 486 590 693 704 721 736 832 928 972

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 4

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 22

> which(myresult2$bootstrap_p<=0.05)

[1] 11 223 251 321 407 415 447 491 567 583 611 688 719 722 723 783 816 826 917
[20] 929 966 968

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 58

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 46

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 61

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 6 60 103 113 139 158 170 173 246 250 253 271 286 296 336 356 361 380 382
[20] 384 386 422 425 434 438 484 493 507 512 527 534 591 606 629 658 669 685 690
[39] 695 706 716 736 747 765 771 806 810 811 824 833 848 861 896 897 911 913 929
[58] 933

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 6 60 103 113 139 154 158 250 253 271 286 296 356 361 382 425 438 444 483
[20] 484 493 507 512 578 591 606 629 658 685 690 706 716 736 765 771 806 810 822
[39] 824 848 896 897 904 911 913 933

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 6 58 60 103 113 139 158 170 173 216 250 253 271 286 296 336 356 361 381
[20] 382 384 386 425 438 475 483 484 493 504 507 512 527 544 578 591 606 629 658
[39] 685 690 695 706 716 736 737 747 765 771 787 806 810 824 848 896 897 911 913
[58] 929 933 993 998

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 14

> con2_adj_p <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adj_p<=0.05/3)

[1] 6

> con3_adj_p <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adj_p<=0.05/3)

[1] 11

> which(con2_adj_p<=0.05/3)

[1] 286 361 484 591 658 765

> which(con3_adj_p<=0.05/3)

[1] 103 139 250 286 361 484 591 629 685 736 765

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000  -none- numeric
ordfit_pvalue 3000  -none- numeric
ordfit_beta1  3000  -none- numeric
permutation_p 3000  -none- numeric
bootstrap_p   3000  -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 49

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 39

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 43

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 3 7 32 69 84 100 103 105 110 123 139 146 170 191 201 228 245 247 275
[20] 315 330 336 371 425 426 439 466 491 496 510 535 537 545 546 603 610 614 654
[39] 693 737 750 779 796 811 831 922 935 943 984

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 3 7 84 100 103 110 123 170 191 193 201 228 245 298 315 330 336 388 425
[20] 426 439 466 478 510 535 537 545 546 557 603 614 654 693 737 811 831 935 943
[39] 984

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 3 7 69 84 99 100 103 110 123 146 170 191 201 228 245 261 275 315 330
[20] 336 341 369 376 388 425 426 466 510 537 545 546 614 693 737 741 750 754 796
[39] 811 831 935 943 984

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 7

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 4

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 9

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "/private/var/folders/db/4tvgx8jx4z3fm1gzlnlzw9rc0000gq/T/Rtmp6jB167/Rinstadd37bc0fed6/RBM/d
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 45
```

```
> sum(diff_results$permutation_p<=0.05)
```

```
[1] 62
```

```
> sum(diff_results$bootstrap_p<=0.05)
```

```
[1] 53
```

```
> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
```

```
> sum(ordfit_adj_p<=0.05)
```

```
[1] 0
```

```
> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
```

```
> sum(perm_adj_p<=0.05)
```

```
[1] 10
```

```
> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
```

```
> sum(boot_adj_p<=0.05)
```

```
[1] 2
```

```
> diff_list_perm <- which(perm_adj_p<=0.05)
```

```
> diff_list_boot <- which(boot_adj_p<=0.05)
```

```
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])
```

```
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
259	cg00234961	0.04192170	0.04321576	0.05707140	0.05327565
627	cg00612467	0.04777553	0.03783457	0.05380982	0.05582291
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
19	0.80831380	0.73306440	0.82968340	0.84917800	
103	0.67349260	0.73510200	0.75715920	0.78981220	
245	0.04208405	0.05284988	0.03775905	0.03955271	
259	0.04030003	0.03996053	0.05086962	0.05445672	
627	0.04740551	0.05332965	0.05775211	0.05579710	
764	0.90575890	0.88760470	0.90756300	0.90946790	
848	0.05230160	0.06119713	0.06542751	0.06240686	
851	0.50820240	0.34657470	0.66276570	0.64634510	
911	0.08633986	0.06765189	0.09070268	0.12417730	

```
928 0.04490690 0.04204062 0.05050039 0.05268215
```

```
diff_results$ordfit_t[diff_list_perm]
19 -2.446404
103 -2.268711
245 1.962457
259 -4.052697
627 -2.239498
764 -1.808081
848 -2.314412
851 -2.841244
911 -3.621731
928 -2.716443
```

```
diff_results$permutation_p[diff_list_perm]
19 0
103 0
245 0
259 0
627 0
764 0
848 0
851 0
911 0
928 0
```

```
> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)
```

```
IlmnID Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
285 cg00263760 0.09050395 0.10197760 0.1480171 0.1224240
833 cg00814580 0.09348613 0.09619816 0.1201044 0.1153424
exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
285 0.1169360 0.1065043 0.1228116 0.1231043
833 0.0957704 0.1159885 0.1286089 0.1411120
diff_results$ordfit_t[diff_list_boot]
285 -3.093997
833 -3.428319
diff_results$bootstrap_p[diff_list_boot]
285 0
833 0
```