

# Package ‘ccmap’

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**Type** Package

**Title** Combination Connectivity Mapping

**Version** 1.0.0

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**Description** Finds drugs and drug combinations that are predicted to reverse or mimic gene expression signatures. These drugs might reverse diseases or mimic healthy lifestyles.

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**LazyData** TRUE

**RoxygenNote** 5.0.1

**VignetteBuilder** knitr

**Suggests** crossmeta, knitr, rmarkdown, testthat, lydata,

**Imports** AnnotationDbi (>= 1.34.4), BiocInstaller, ccddata (>= 0.99.4), doParallel (>= 1.0.10), data.table (>= 1.9.6), foreach (>= 1.4.3), parallel (>= 3.3.1), xgboost (>= 0.4.4)

**biocViews** GeneExpression, Transcription, Microarray, DifferentialExpression

**NeedsCompilation** no

## R topics documented:

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`get_dprimes`*Extract unbiased effect sizes from meta-analysis by crossmeta.*

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**Description**

Function extracts mu (overall mean effect size) and dprimes (unbiased effect sizes from each contrast).

**Usage**

```
get_dprimes(es)
```

**Arguments**

`es`                      Result of call to `es_meta`.

**Details**

Result used to query connectivity map drugs and predicted drug combinations.

**Value**

List containing:

`meta`                      Named numeric vector with overall mean effect sizes for all genes from meta-analysis.

`contrasts`                List of named numeric vectors (one per contrast) with unbiased effect sizes for all measured genes.

**See Also**

[es\\_meta](#).

**Examples**

```
library(crossmeta)
library(lydata)

data_dir <- system.file("extdata", package = "lydata")

# gather GSE names
gse_names <- c("GSE9601", "GSE15069", "GSE50841", "GSE34817", "GSE29689")

# load previous differential expression analysis
anals <- load_diff(gse_names, data_dir)

# run meta-analysis
es <- es_meta(anals)

#get dprimes
dprimes <- get_dprimes(es)
```

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query_combos	<i>Get overlap between query and predicted drug combination signatures.</i>
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### Description

Drugs with the largest positive and negative net overlap are predicted to, respectively, mimic and reverse the query signature. A value of 1 would indicate that all drug and query genes are regulated in the same direction and have the same order when sorted by absolute changes in differential expression. A value of -1 would indicate that all drug and query genes are regulated in the opposite direction and have the same order when sorted by absolute changes in differential expression.

### Usage

```
query_combos(query_genes, method = "average", include = NULL,
             ncores = parallel::detectCores())
```

### Arguments

query_genes	Named numeric vector of differential expression values for query genes. Usually 'meta' slot of get_dprimes result.
method	One of 'average' (default) or 'ml' (machine learning - see details and vignette).
include	Character vector of cmap drug names for which combinations with all other cmap drugs will be predicted and queried. If NULL (default), all 856086 two drug combinations will be predicted and queried.
ncores	Integer, number of cores to use for method 'average'. Default is to use all cores.

### Details

To predict and query all 856086 two-drug combinations, the 'average' method can take as little as 10 minutes (Intel Core i7-6700). The 'ml' (machine learning) method takes two hours on the same hardware and requires ~10GB of RAM but is slightly more accurate. Both methods will run faster by specifying only a subset of drugs using the include parameter. To speed up the 'ml' method, the MRO+MKL distribution of R can help substantially ([link](#)).

### Value

Vector of numeric values between 1 and -1 indicating extent of overlap between query and drug combination signatures (see description).

### Examples

```
library(lydata)
library(crossmeta)

# location of data
data_dir <- system.file("extdata", package = "lydata")

# gather GSE names
gse_names <- c("GSE9601", "GSE15069", "GSE50841", "GSE34817", "GSE29689")

# load previous analysis
```

```

anals <- load_diff(gse_names, data_dir)

# perform meta-analysis
es <- es_meta(anals)

# get dprimes
dprimes <- get_dprimes(es)

# query combinations of metformin and all other cmap drugs
top_met_combos <- query_combos(dprimes$meta, include = 'metformin', ncores = 1)

# previous query but with machine learning method
# top_met_combos <- query_combos(dprimes$meta, 'ml', 'metformin')

# query all cmap drug combinations
# top_combos <- query_combos(dprimes$meta)

# query all cmap drug combinations with machine learning method
# top_combos <- query_combos(dprimes$meta, 'ml')

```

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query\_drugs

*Get overlap between query and drug signatures.*


---

### Description

Determines the volume under the surface formed by plotting net overlap (z) as a function of number of drug and query genes (x and y).

### Usage

```
query_drugs(query_genes, drug_info = NULL, sorted = TRUE)
```

### Arguments

query_genes	Named numeric vector of differential expression values for query genes. Usually 'meta' slot of get_dprimes result.
drug_info	Matrix of differential expression values for drugs or drug combinations. Rows are genes, columns are drugs.
sorted	Would you like the results sorted in decreasing order of overlap? Default is TRUE.

### Details

Drugs with the largest positive and negative net overlap are predicted to, respectively, mimic and reverse the query signature. A value of 1 would indicate that all drug and query genes are regulated in the same direction and have the same order when sorted by absolute changes in differential expression. A value of -1 would indicate that all drug and query genes are regulated in the opposite direction and have the same order when sorted by absolute changes in differential expression.

### Value

Vector of numeric values between 1 and -1 indicating extent of overlap between query and drug signatures (see description).

**See Also**

[query\\_combos](#) to get overlap between query and predicted drug combination signatures.

**Examples**

```
# create drug signatures
genes <- paste("GENE", 1:1000, sep = "_")
set.seed(0)

drug_info <- data.frame(row.names = genes,
                        drug1 = rnorm(1000, sd = 2),
                        drug2 = rnorm(1000, sd = 2),
                        drug3 = rnorm(1000, sd = 2))

# query signature is drug3
query_sig <- drug_info$drug3
names(query_sig) <- genes

res <- query_drugs(query_sig, as.matrix(drug_info))
```

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sum\_rowcolCumsum

*Sum of cumulative sum computed over rows then columns of matrix.*

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**Description**

Equivalent to computing the cumulative sum of a matrix over rows, then over columns, then summing every value (though much faster and more memory efficient).

**Usage**

```
sum_rowcolCumsum(x, i, j)
```

**Arguments**

x                    Numeric vector of non-zero values of matrix.  
i                    Integer vector of row indices of x.  
j                    Integer vector of column indices of x.

**Value**

Numeric value equal to the sum of the cumulative sum computed over rows then columns of a matrix.

**Examples**

```
x <- c(1, 1, 1, -1) # non-zero values of matrix
i <- c(1, 2, 3, 4) # row indices of x
j <- c(4, 1, 3, 2) # col indices of x

sum_rowcolCumsum(x, i, j)
```

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