

Package ‘mAPKL’

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Type Package

Title A Hybrid Feature Selection method for gene expression data

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Description We propose a hybrid FS method (mAP-KL), which combines multiple hypothesis testing and affinity propagation (AP)-clustering algorithm along with the Krzanowski & Lai cluster quality index, to select a small yet informative subset of genes.

License GPL (>= 2)

biocViews FeatureExtraction, DifferentialExpression, Microarray,
GeneExpression

NeedsCompilation no

R topics documented:

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mAPKL-package	<i>A hybrid feature selection method for gene expression data</i>
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Description

A hybrid FS method (mAP-KL), which combines multiple hypothesis testing and affinity propagation (AP) clustering algorithm along with the Krzanowski & Lai cluster quality index, to select an informative subset of genes.

Details

Package: mAPKL
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Author(s)

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References

A. Sakellariou, D. Sanoudou, and G. Spyrou, "Combining multiple hypothesis testing and affinity propagation clustering leads to accurate, robust and sample size independent classification on gene expression data, " BMC Bioinformatics, vol. 13, p. 270, 2012.

Annot-class	<i>Class "Annot"</i>
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Description

S4 class for storing Annot analysis results

Slots

The following slots are defined for [Annot](#) objects:

- results: The accumulated annotation results
- probe: The probe id
- symbol: The official gene symbol
- entrezId: The Entrez gene Identifier
- ensemblId: The ensembl ID as indicated by ensembl
- map: The cytoband locations of the gene

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

annotate *Genome annotation of the "exemplars".*

Description

The user may extract several genome specific information for the "exemplars" as described in the microarray annotation file.

Usage

annotate(exemplars,chip)

Arguments

- exemplars The "exemplars" of the mAPKL class.
- chip The platforms's name of the microarray chip used (e.g. "hgu133plus2.db")

Details

This function uses as key the probe id and returns the mathing information as described in the gene chip annotation file. The returned information are usually multiple to the number of probe ids (one to many relationship).

Value

- results The accumulated annotation results.
- probe The probe id.
- symbol The official gene symbol.
- entrezId The Entrez gene Identifier.
- ensemblId The ensembl ID as indicated by ensembl.
- map The cytoband locations of the gene.

Author(s)

Argiris Sakellariou

Examples

```
## We use the "exemplars" from the mAPKL.Rd example

exemplrs <- c(24, 26, 42, 45, 63, 81, 95, 99, 102, 113, 134, 135, 145, 152, 168)
names(exemplrs) <- c("215717_s_at", "1561358_at", "222752_s_at", "233922_at",
"218871_x_at", "33323_r_at", "244311_at", "220932_at", "205508_at", "209596_at",
"215180_at", "1560638_a_at", "201852_x_at", "229947_at", "221731_x_at")

gene.info <- annotate(exemplrs, "hgu133plus2.db")
```

classification

*Classify samples according to the SVM algorithm***Description**

This function performs classification through the Support Vector Machines (SVM) algorithm. The algorithm applies on the "exemplars" dataset. It produces a classification result either on the training set or on a validation set. This function estimates how well the selected "genes" from mAP-KL method discriminate between two phenotypes. The default SVM settings are: "linear" kernel and 5-folds cross-validation. Regarding the parameters for the "linear" kernel, cost parameter, and for the "radial" kernel, cost and gamma parameters, are estimated automatically through the `tune.svm` function as described in `e1071` r-package.

Usage

```
classification(trExemplObj, classLabels, valExemplObj=NULL, kf=5, kernel="linear")
```

Arguments

<code>trExemplObj</code>	The exemplars train eSet object.
<code>classLabels</code>	The varLabels name in the eSet object where the class labels are stored e.g "type".
<code>valExemplObj</code>	The exemplars validation eSet object (if not NULL).
<code>kf</code>	The k-folds value of the cross-validation parameter. The default value is 5-folds. By setting "Loo" or "LOO" a Leave-One-Out Cross Validation is performed
<code>kernel</code>	The type of kernel used for the classification analysis. The default kernel is "linear"

Value

<code>classL</code>	The labels of the train set
<code>valClassL</code>	The labels of the validation set if not NULL
<code>predLbls</code>	The predicted labels according to the classification analysis

Author(s)

Argiris Sakellariou

Examples

```
library(mAPKLData)
data(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(breast$trainData)
normTestData <- preprocess(breast$testData)

exprs(breast$trainData)<-normTrainData$c1L2.normdata
exprs(breast$testData)<-normTestData$c1L2.normdata

out.c1L2 <- mAPKL(trObj=breast$trainData, classLabels="type",
valObj=breast$testData,dataType=7)

clasPred <- classification(trExemplObj=out.c1L2@exemplTrain, classLabels="type",
valExemplObj=out.c1L2@exemplTest)
```

Classify-class

Class "Classify"

Description

S4 class for storing Classify analysis results

Slots

The following slots are defined for [Classify](#) objects:

classL: Number of samples in the data set

valClassL: Subset of samples used for leveraged clustering

predLbls: Vector containing indices of exemplars

AUC: The Area Under the ROC curve as a degree of samples discrimination

Accuracy: The classification accuracy

MCC: The MCC classification measure

Specificity: The degree of true negative's identification

Sensitivity: The degree of true positive's identification

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

DataLD-class	<i>Class "DataLD"</i>
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Description

S4 class for storing DataLD analysis results

Slots

The following slots are defined for [DataLD](#) objects:

trainObj: An eSet class object of a training set

valObj: An eSet class object of a validation set

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

loadFiles	<i>Imports gene expression data</i>
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Description

This function loads the train set, the class labels files as well as the test or validation file if any. Then we may perform normalization and (or) log2 transformation.

Usage

```
loadFiles(filesPath, trainFile, labelsFile, validationFile=NULL,
validationLabels=NULL)
```

Arguments

filesPath	The path where the files are stored
trainFile	The genes and the relevant intensity values for feature selection analysis. The file should be of tab-delimited format
labelsFile	The class labels of the samples
validationFile	A further file with genes and intensity values used for validation purposes
validationLabels	The class labels of the validation samples

Value

An object of Class [DataLD](#)

Author(s)

Argiris Sakellariou

Description

We first employ the `mt.maxT` function from the "multtest" package to rank the genes of the training set and then we reserve the top N genes e.g.(N = 200) for further exploitation. Prior to clustering analysis with Affinity Propagation we apply the index of Krzanowski and Lai as included in the "ClusterSim" package to determine the number of clusters solely on the disease samples of the training test set. The final step involves the cluster analysis with the AP clustering method as in the "apcluster" package, which detects n (n = k, the Krzanowski and Lai index) clusters among the top N genes and provides us with a list of the most representative genes of each cluster, the so called exemplars.

Usage

```
mAPKL(trObj,classLabels,valObj=NULL,dataType=6,statTest="t",permutations=1000,
features=200,minClusters=2,maxClusters=50,FC="limma",bimaxit=50,r=2)
```

Arguments

<code>trObj</code>	The train eSet object.
<code>classLabels</code>	The varLabels name in the eSet object where the class labels are stored e.g "type".
<code>valObj</code>	The validation eSet object (if not NULL).
<code>dataType</code>	The type of the data e.g 6-ratio data without normalization and 7-interval or mixed (ratio & interval) data without normalization as described in "clusterSim" package.
<code>statTest</code>	The statistical test applied to the geneIntensities. The available tests described in <code>mt.maxT</code> documentation in "multtest" package.
<code>permutations</code>	The number of permutations.
<code>features</code>	The top N genes to be kept.
<code>minClusters</code>	The minimum number of clusters that can be identified.
<code>maxClusters</code>	The maximum number of clusters that can be identified.
<code>FC</code>	The Fold Change of the exemplars according to "Limma" (default). Alternatively the "SAM" approach may be computed.
<code>bimaxit</code>	The maximum number of bisection steps performed by the AP algorithm. The (default) value is "50".
<code>r</code>	The argument r is used to transform the resulting distances by computing the r-th power. To obtain negative squared distances as in Frey's and Dueck's (use r=2 as default).

Value

rankedIntens	The top N ranked genes with their intensity values
exemplTrain	The intensity values of the exemplars in the training set
exemplTest	The intensity values of the exemplars in the validation set if not NULL
statistic	A list with the overall results of the "mt.maxT" analysis
adjp	The adjusted p-values according to the statistical analysis
pVal	The raw p-values according to the statistical analysis
fc	The Fold Change of the exemplars
exemplars	The selected "significant" probe ids/genes
clusters	The probe ids/genes per cluster

Author(s)

Argiris Sakellariou

References

A. Sakellariou, D. Sanoudou, and G. Spyrou, "Combining multiple hypothesis testing and affinity propagation clustering leads to accurate, robust and sample size independent classification on gene expression data," *BMC Bioinformatics*, vol. 13, p. 270, 2012.

Examples

```
## Using separate train-test samples
## Load the necessary files based on Breast cancer data as included in the
## package mAPKLData

library(mAPKLData)
data(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(breast$trainData)
normTestData <- preprocess(breast$testData)

exprs(breast$trainData) <- normTrainData$cLL2.normdata
exprs(breast$testData) <- normTestData$cLL2.normdata

out.cLL2 <- mAPKL(trObj=breast$trainData, classLabels="type",
valObj=breast$testData, dataType=7)
```

mAPKLRes-class	Class "mAPKLRes"
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Description

S4 class for storing mAPKL analysis results

Slots

The following slots are defined for `mAPKLRes` objects:

`rankedIntens`: The top N ranked genes along with their intensity values

`exemplTrain`: An `eSet` class object formed with the exemplars of the training set

`exemplTest`: An `eSet` class object formed with the exemplars of the validation set if not NULL

`statistic`: A list with the overall results of the "mt.maxT" analysis

`adjp`: The adjusted p-values according to the statistical analysis

`pVal`: The raw p-values according to the statistical analysis

`fc`: The Fold Change of the exemplars

`exemplars`: The selected "significant" probe ids/genes

`clusters`: The probe ids/genes per cluster

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

metrics	Computes several clasification metrics
---------	--

Description

This function calculates several classification related metrics. It uses the original and the predicted samples' labels to quantify the quality of the classification process. Those meassures give us a direct outlook of the selected "genes" and how well discriminate between two phenotypes.

Usage

```
metrics(classLbIs, predLbIs)
```

Arguments

`classLbIs` The initial class labels.

`predLbIs` The predicted class labels.

Value

AUC	The Area Under the ROC curve as a degree of samples discrimination
Accuracy	The classification accuracy
MCC	The MCC classification measure
Specificity	The degree of true negative's identification
Sensitivity	The degree of true positive's identification

Author(s)

Argiris Sakellariou

Examples

```
## Suppose 'val' represent the correct validation set labels
## and 'predictions' the predicted labels according to an SVM model

val <- c(rep(0,8),rep(1,4))
predictions <- c(rep(0,6),1,1,rep(1,3),0)
perfMetrics <- metrics(classLbIs=val, predLbIs=predictions)
```

NetAttr-class

Class "NetAttr"

Description

S4 class for storing some network characteristics of the top ranked genes

Slots

The following slots are defined for [NetAttr](#) objects:

edgelist: The Node1 & Node2 & Weight edgelist matrix

degree: The Local, Global and their Weighted values of the "degree" characteristic

closeness: The Weighted values of the Local and Global "closeness" characteristic

betweenness: The Weighted values of the Local and Global "betweenness" characteristic

transitivity: The Weighted values of the Local and Global "transitivity" characteristic

Author(s)

Argiris Sakellariou <a.sakellariou@gonkhosp.gr>

netwAttr	<i>Calculates network characteristics</i>
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Description

Calculate some basic network characteristics of the top ranked genes

Usage

```
netwAttr(mAPKLobj, net="clr")
```

Arguments

mAPKLobj	An object of mAPKL class.
net	The network reconstruction method to be employed. The user may select between "clr" (default), "aracne.a" and "aracne.m".

Details

It calculates some basic network characteristics. Those include the "degree", the "closeness", the "betweenness", and finally the "transitivity" or else clustering coefficient. We calculate the weighted values for both local and global scores.

The three available network reconstruction options are:

clr: Context Likelihood or Relatedness Network

aracne.a: Algorithm for the Reconstruction of Accurate Cellular Networks (additive model)

aracne.m: Algorithm for the Reconstruction of Accurate Cellular Networks (multiplicative model)

Value

Upon successful completion, the function returns an [NetAttr](#) object.

Author(s)

Argiris Sakellariou

Examples

```
library(mAPKLData)
data(mAPKLData)
breast <- sampling(Data=mAPKLData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(breast$trainData)
normTestData <- preprocess(breast$testData)

exprs(breast$trainData) <- normTrainData$cLL2.normdata
exprs(breast$testData) <- normTestData$cLL2.normdata

out.cLL2 <- mAPKL(trObj=breast$trainData, classLabels="type",
```

```
valObj=breast$testData, dataType=7)
net.attr <- netwAttr(mAPKLObj=out.c1L2)
```

preprocess	<i>Performs normalization and/or log2 transformation</i>
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Description

This function performs normalization and/or log2 transformation on gene expression data.

Usage

```
preprocess(exprsObj, log2=TRUE, norm="ALL", destname=NULL)
```

Arguments

exprsObj	An eSet object where its assay data will be normalized
log2	Performs logarithmic transformation of base 2 prior to any normalization. The default value is TRUE
norm	The user may define a specific normalization method rather than "ALL" which is the default case. The available abbreviations are described in the details section
destname	Here we define the destination path and the name of the jpeg file with the density plots. The default path is the working directory

Details

The available normalization methods are:

Mean-centering normalization "mc"

z-score normalization "z"

Quantile normalization "q"

Cyclic loess normalization "cl"

Mean-centering normalization and log2 transformation "mcL2"

z-score normalization and log2 transformation "zL2"

Quantile normalization and log2 transformation "qL2"

Cyclic loess normalization and log2 transformation "clL2"

Value

rawdata	The initial gene expression values
mc.normdata	The values after 'mean-centering' normalization
z.normdata	The values after 'z-score' normalization
q.normdata	The values after 'quantile' normalization
cl.normdata	The values after 'cyclic loess' normalization

mCL2.normdata	The values after 'mean-centering' normalization and log2
zL2.normdata	The values after 'z-score' normalization and log2
qL2.normdata	The values after 'quantile' normalization and log2
c1L2.normdata	The values after 'cyclic loess' normalization and log2

Author(s)

Argiris Sakellariou

Examples

```
library(mAPKLDData)
data(mAPKLDData)
varLabels(mAPKLDData)
breast <- sampling(Data=mAPKLDData, valPercent=40, classLabels="type", seed=135)
normTrainData <- preprocess(exprsObj=breast$trainData)
```

probes2pathways

Extract pathways from "exemplars"

Description

The user may extract the pathways where the "exemplars" are involved with the aid of the reactome database.

Usage

```
probes2pathways(annotObj)
```

Arguments

annotObj The "Annot" class object.

Details

This function utilizes the "Annot" class object as returned by the "annotate" function to extract the pathways where the "exemplars" are involved with the aid of the "reactome" database. We employ the probe Ids for the matching.

Author(s)

Argiris Sakellariou

Examples

```
## We use the "exemplars" from the mAPKL.Rd example

exemplrs <- c(24, 26, 42, 45, 63, 81, 95, 99, 102, 113, 134, 135, 145, 152, 168)
names(exemplrs) <- c("215717_s_at", "1561358_at", "222752_s_at", "233922_at",
"218871_x_at", "33323_r_at", "244311_at", "220932_at", "205508_at", "209596_at",
"215180_at", "1560638_a_at", "201852_x_at", "229947_at", "221731_x_at")

gene.info <- annotate(exemplrs, "hgu133plus2.db")

## We now use the "gene.info" to identify the relevant pathways

probes2pathways(gene.info)
```

report

Produce an HTML report of the mAP-KL analysis

Description

This function gathers the results of several analysis sessions (feature selection, classification, annotation and network) and produces a report in HTML format.

Usage

```
report(mAPKLObj, ClassifyObj, AnnotObj=NULL, netObj=NULL, file)
```

Arguments

mAPKLObj	An object of mAPKL class.
ClassifyObj	An object of mAPKL class.
AnnotObj	An object of Annot class.
netObj	An object of NetAttr class.
file	The full path and the name of the produced report

Details

It presents the data samples with their phenotype labels, the exemplars with their statistical scores (adj.p-value, p-value and fc), and network characteristics (like weighted local degree, closeness, betweenness, transitivity) if such a network analysis has been performed. In addition, the report presents the classification performance achieved by those exemplars (either cross-validation or hold-out), and finally several annotation information (symbol, entrez id, ensemble id, and their chromosomal location) if an annotation analysis has been carried out.

Value

Upon successful completion an HTML report is saved in the working directory.

Author(s)

Argiris Sakellariou

Examples

```
## When a network attributes object is present
## Not run: report(out,class.pred,class.metrics,netObj=net.attr,
"C:/.../.../mAPKLreport.html")#Define a local path to store your report
## End(Not run)

## When an annotation object is present
## Not run: report(out,class.pred,class.metrics,gene.info,
"C:/.../.../mAPKLreport.html")#Define a local path to store your report
## End(Not run)

## Without annotation and network attributes objects
## Not run: report(out,class.pred,class.metrics,
file="C:/.../.../mAPKLreport.html")#Define a local path to store your report
## End(Not run)
```

sampling

*Splits a dataset to a train and a test sets of a user defined percentage***Description**

This function takes as input a dataset and splits it into a train and a test set based to a user defined percentage.

Usage

```
sampling(Data,valPercent,classLabels,seed)
```

Arguments

Data	The input dataset to be split as an eSet object.
valPercent	The percentage of the input dataset used for validation purposes e.g. 40.
classLabels	The varLabels name in the eSet object where the class labels are stored e.g "type".
seed	Setting the seed number for reproducible sampling. The default value is 1.

Value

trainData	The data used for training as an eSet object
testData	The data used for validation as an eSet object

Author(s)

Argiris Sakellariou

Examples

```
library(mAPKLDData)
data(mAPKLDData)
breast <- sampling(Data=mAPKLDData, valPercent=40, classLabels="type", seed=135)
```


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