

Introduction to the pageRank Package

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1 Introduction

1.1 Background

The *pageRank* package provides implementations of temporal PageRank as defined by [1], as well as multiplex PageRank as defined by [2]. As the extension of original steady-state PageRank [3,4] in temporal networks, temporal PageRank ranks nodes based on their connections that change over time. Multiplex PageRank, on the other hand, extends PageRank analysis to multiplex networks. In such networks, the same nodes might interact with one another in different layers. Multiplex PageRank is calculated

according to the topology of a predefined base network, with regular PageRank of other supplemental networks as edge weights and personalization vector.

PageRank-related approaches can be applied to prioritize key transcriptional factors (TFs) in gene regulatory networks (GRNs). Specifically, the *pageRank* package provides functions for generating temporal GRNs from corresponding static counterparts. The *pageRank* package also provides functions for converting multi-omics, e.g. gene expression, chromatin accessibility and chromosome conformation profiles to multiplex GRNs. Such temporal and multiplex GRNs can thus be used for temporal and multiplex PageRank-based TF prioritization, respectively.

1.2 Installation

pageRank requires the R version 4.0 or later, packages *BSgenome.Hsapiens.UCSC.hg19*, *TxDb.Hsapiens.UCSC.hg19.knownGene*, *org.Hs.eg.db*, *annotate*, *GenomicFeatures*, *JASPAR2018*, *TFBSTools* and *bcellViper*, to run the examples. After installing R, all required components can be obtained with:

```
if (!requireNamespace("BiocManager", quietly=TRUE)) install.packages("BiocManager")
BiocManager::install("BSgenome.Hsapiens.UCSC.hg19")
BiocManager::install("TxDb.Hsapiens.UCSC.hg19.knownGene")
BiocManager::install("org.Hs.eg.db")
BiocManager::install("annotate")
BiocManager::install("GenomicFeatures")
BiocManager::install("JASPAR2018")
BiocManager::install("TFBSTools")
BiocManager::install("bcellViper")
```

2 PageRank Analysis

2.1 Temporal PageRank

We applied `diff_graph()` to calculate temporal PageRank. This is a simplified version of temporal PageRank described by [1] by only analyzing temporally adjacent graph pairs.

```
> library(pageRank)
> set.seed(1)
> graph1 <- igraph::erdos.renyi.game(100, 0.01, directed = TRUE)
> igraph::V(graph1)$name <- 1:100
> #the 1st graph with name as vertex attributes
> set.seed(2)
> graph2 <- igraph::erdos.renyi.game(100, 0.01, directed = TRUE)
```

```

> igraph::V(graph2)$name <- 1:100
> #the 2nd graph with name as vertex attributes
> diff_graph(graph1, graph2)

IGRAPH 2152967 DN-- 98 190 --
+ attr: name (v/c), pagerank (v/n), moi (e/n)
+ edges from 2152967 (vertex names):
 [1] 1 ->60 2 ->15 2 ->57 3 ->10 3 ->16 3 ->84 4 ->43 5 ->20 5 ->66
[10] 5 ->72 5 ->81 5 ->91 6 ->25 6 ->50 7 ->37 7 ->67 7 ->73 7 ->85
[19] 8 ->80 9 ->90 10->26 11->6 11->100 12->70 12->82 12->92 13->30
[28] 13->48 13->51 13->61 15->74 15->77 16->85 17->31 17->32 17->34
[37] 17->50 17->58 19->17 19->96 20->23 20->79 20->87 21->41 21->45
[46] 22->4 22->41 23->57 24->61 25->66 26->34 26->39 26->72 27->22
[55] 27->43 28->98 29->95 30->84 32->49 33->10 34->16 34->99 35->81
[64] 36->17 36->33 36->45 36->53 36->77 37->33 37->54 38->6 38->17
+ ... omitted several edges

```

Differential graph graph1-graph2 will be outputted. The Differential graph has "moi (mode of interaction, 1 and -1 for interactions gained and losed in graph1, respectively)" as edge attribute. The Differential graph has "pagerank" and "name" as vertex attributes.

2.2 Multiplex PageRank

We applied `multiplex_page_rank()` to calculate multiplex PageRank following defination by [2].

```

> set.seed(1)
> graph1 <- igraph::erdos.renyi.game(100, 0.01, directed = TRUE)
> igraph::V(graph1)$name <- 1:100
> igraph::V(graph1)$pagerank <- igraph::page_rank(graph1)$vector
> #the base graph with pagerank and name as vertex attributes.
> set.seed(2)
> graph2 <- igraph::erdos.renyi.game(100, 0.01, directed = TRUE)
> igraph::V(graph2)$name <- 1:100
> igraph::V(graph2)$pagerank <- igraph::page_rank(graph2)$vector
> #the supplemental graph with pagerank and name as vertex attributes.
> multiplex_page_rank(graph1, graph2)

```

	1	2	3	4	5	6
0.022431793	0.004376548	0.005101288	0.021806420	0.004038261	0.014009056	
	7	8	9	10	11	12
0.017033776	0.004038261	0.010897818	0.022024129	0.004376548	0.004038261	
	13	14	15	16	17	18

0.004038261	0.004038261	0.011984174	0.010662594	0.012861595	0.004038261
19	20	21	22	23	24
0.004038261	0.006973192	0.007883792	0.018708510	0.009449760	0.008580800
25	26	27	28	29	30
0.009992110	0.023175991	0.016861132	0.007081820	0.004038261	0.007696307
31	32	33	34	35	36
0.017884438	0.004465826	0.019332930	0.005497036	0.026386118	0.014886821
37	38	39	40	41	42
0.004473382	0.012774746	0.008259414	0.008244753	0.010494265	0.015930714
43	44	45	46	47	48
0.017448552	0.007901711	0.008117970	0.007680122	0.004038261	0.004263786
49	50	51	52	53	54
0.011679744	0.020924465	0.008291128	0.009090471	0.008316500	0.008569083
55	56	57	58	59	60
0.004038261	0.011492897	0.013013291	0.006154228	0.009112068	0.005053121
61	62	63	64	65	66
0.011557465	0.008125871	0.004038261	0.009595523	0.004038261	0.013377737
67	68	69	70	71	72
0.004473382	0.005786216	0.006188052	0.011348562	0.004807863	0.021639449
73	74	75	76	77	78
0.005681911	0.011938368	0.004664091	0.028329952	0.005204011	0.008601557
79	80	81	82	83	84
0.007514945	0.004714834	0.037833945	0.005251514	0.004376548	0.007672082
85	86	87	88	89	90
0.017889151	0.004376548	0.013828925	0.005346400	0.004038261	0.010681597
91	92	93	94	95	96
0.011139970	0.009048308	0.015566866	0.009116546	0.006681434	0.004714834
97	98	99	100		
0.012194456	0.008225016	0.005368584	0.004931364		

Multiplex PageRank values corresponded to nodes in graph1 (base network) will be outputed.

2.3 Adjusting PageRank Calculations

The `clean_graph()` can remove nodes by residing subgraph sizes, vertex names and PageRank values. We thus can adjust graphs for PageRank calculation.

```
> set.seed(1)
> graph <- igraph::erdos.renyi.game(100, 0.01, directed = TRUE)
> igraph::V(graph)$name <- 1:100
> igraph::V(graph)$pagerank <- igraph::page_rank(graph)$vector
```

```
> #the graph to be cleaned, with pagerank and name as vertex attributes.
> clean_graph(graph, size=5)
```

```
IGRAPH de34ea0 DN-- 82 96 -- Erdos renyi (gnp) graph
+ attr: name (g/c), type (g/c), loops (g/l), p (g/n), name (v/n),
| pagerank (v/n)
+ edges from de34ea0 (vertex names):
 [1] 72-> 1 88-> 3 22-> 4 11-> 6 65-> 6 87-> 6 60-> 7 85-> 7
 [9] 84-> 9 33-> 10 100-> 10 11->100 2-> 15 40-> 15 3-> 16 34-> 16
[17] 19-> 17 46-> 17 5-> 20 69-> 20 100-> 20 92-> 21 27-> 22 83-> 23
[25] 42-> 24 6-> 25 10-> 26 74-> 27 94-> 27 43-> 31 36-> 33 38-> 35
[33] 59-> 35 90-> 35 60-> 36 70-> 36 53-> 38 26-> 39 46-> 40 88-> 40
[41] 21-> 41 71-> 41 49-> 42 65-> 42 77-> 42 87-> 43 100-> 43 52-> 44
[49] 21-> 45 54-> 46 32-> 49 92-> 49 6-> 50 17-> 50 43-> 52 54-> 52
+ ... omitted several edges
```

Adjusted graph will be outputted, with "pagerank" and "name" as vertex attributes. The `adjust_graph()` can re-calculate PageRank with updated damping factor, personalized vector and edge weights.

```
> set.seed(1)
> graph <- igraph::erdos.renyi.game(100, 0.01, directed = TRUE)
> igraph::V(graph)$name <- 1:100
> igraph::V(graph)$pagerank <- igraph::page_rank(graph, damping=0.85)$vector
> #the graph to be adjusted, with pagerank and name as vertex attributes.
> adjust_graph(graph, damping=0.1)
```

```
IGRAPH ce87d6a DN-- 100 98 -- Erdos renyi (gnp) graph
+ attr: name (g/c), type (g/c), loops (g/l), p (g/n), name (v/n),
| pagerank (v/n)
+ edges from ce87d6a (vertex names):
 [1] 72-> 1 88-> 3 22-> 4 11-> 6 65-> 6 87-> 6 60-> 7 85-> 7
 [9] 84-> 9 33-> 10 100-> 10 11->100 2-> 15 40-> 15 3-> 16 34-> 16
[17] 19-> 17 46-> 17 5-> 20 69-> 20 100-> 20 92-> 21 27-> 22 83-> 23
[25] 42-> 24 6-> 25 10-> 26 74-> 27 94-> 27 63-> 30 43-> 31 36-> 33
[33] 38-> 35 59-> 35 90-> 35 60-> 36 70-> 36 53-> 38 26-> 39 46-> 40
[41] 88-> 40 21-> 41 71-> 41 49-> 42 65-> 42 77-> 42 87-> 43 100-> 43
[49] 52-> 44 21-> 45 54-> 46 32-> 49 92-> 49 6-> 50 17-> 50 13-> 51
+ ... omitted several edges
```

Adjusted graph will be outputted, with updated "pagerank" and "name" as vertex attributes.

Please note `diff_graph()`, `multiplex_page_rank()`, `clean_graph()` and `adjust_graph()` can be used in combination for customized PageRank analysis tasks.

3 Prioritizing TFs in GRNs

3.1 Generating GRNs from Multi-Omics Profiles

The `aracne_network()` can re-format ARACNe network in regulon object for PageRank analysis. It can also handle GRNs reverse engineered using other algorithms, as long as such GRNs are written in regulon object.

```
> library(bcellViper)
> data(bcellViper)
> head(aracne_network(regulon[1:10]))
```

	reg	target	direction
1	AATF	SAMM50	1
2	AATF	DRG1	1
3	AATF	ATIC	1
4	AATF	SMARCC1	1
5	AATF	AHCY	1
6	AATF	HSD17B10	1

The `accessibility_network()` can build network from accessibility, e.g. ATAC-Seq peaks.

```
> table <- data.frame(Chr=c("chr1", "chr1"), Start=c(713689, 856337), End=c(714685, 856337),
+                     row.names=c("A", "B"), stringsAsFactors=FALSE)
> regulators=c("FOXF2", "MZF1")
> #peaks and regulators to be analyzed
>
> library(GenomicRanges)
> library(GenomicFeatures)
> library(TxDb.Hsapiens.UCSC.hg19.knownGene)
> library(org.Hs.eg.db)
> library(annotate)
> promoter <- promoters(genes(TxDb.Hsapiens.UCSC.hg19.knownGene))
> names(promoter) <- getSYMBOL(names(promoter), data="org.Hs.eg")
> promoter <- promoter[!is.na(names(promoter))]
> #get promoter regions
>
> library(JASPAR2018)
> library(TFBSTools)
> library(motifmatchr)
> pfm <- getMatrixSet(JASPAR2018, list(species="Homo sapiens"))
> pfm <- pfm[unlist(lapply(pfm, function(x) name(x))) %in% regulators]
```

```

> #get regulator position frequency matrix (PFM) list
>
> library(BSgenome.Hsapiens.UCSC.hg19)
> accessibility_network(table, promoter, pfm, "BSgenome.Hsapiens.UCSC.hg19")

```

```

      target  reg
1 LOC100288069 FOXF2
2 LOC100288069  MZF1
3   LINC02593 FOXF2
4     SAMD11 FOXF2
5   LINC02593  MZF1
6     SAMD11  MZF1

```

The `conformation_network()` can build network from conformation, e.g. HiChIP records.

```

> table <- data.frame(Chr1=c("chr1", "chr1"), Position1=c(569265, 713603), Strand1=c(
+                               Chr2=c("chr4", "chr1"), Position2=c(206628, 715110), Strand2=c(
+                               row.names=c("A", "B"), stringsAsFactors=FALSE)
> regulators=c("FOXF2", "MZF1")
> #peaks and regulators to be analyzed
>
> promoter <- promoters(genes(TxDb.Hsapiens.UCSC.hg19.knownGene))
> names(promoter) <- getSYMBOL(names(promoter), data="org.Hs.eg")
> promoter <- promoter[!is.na(names(promoter))]
> #get promoter regions
>
> pfm <- getMatrixSet(JASPAR2018, list(species="Homo sapiens"))
> pfm <- pfm[unlist(lapply(pfm, function(x) name(x))) %in% regulators]
> #get regulator position frequency matrix (PFM) list
>
> conformation_network(table, promoter, pfm, "BSgenome.Hsapiens.UCSC.hg19")

```

```

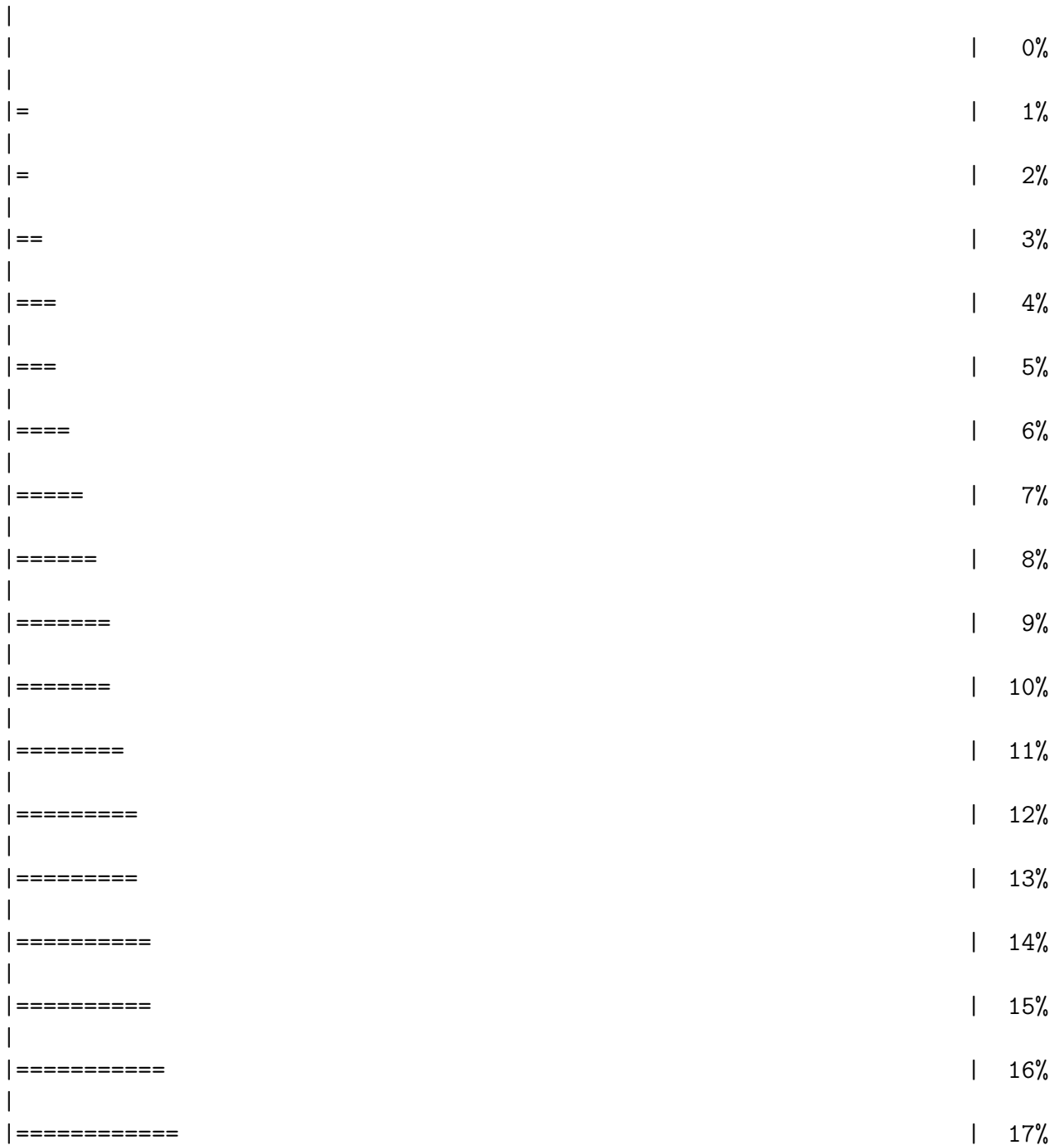
      target  reg
1     ZNF876P  MZF1
2 LOC100288069 FOXF2
3 LOC100288069  MZF1

```

3.2 Filter GRNs with Expression Profiles

The `P_graph()` can filter GRNs by quantifying joint and margin probability distributions of regulator-target pairs. Statistically significant non-random regulator-target pairs will be kept.

```
> dset <- exprs(dset)
> net <- do.call(rbind, lapply(1:10, function(i, regulon){
+   data.frame(reg=rep(names(regulon)[i], 10),
+             target=names(regulon[[i]][[1]])[1:10],
+             stringsAsFactors = FALSE)}, regulon=regulon))
> P_graph(dset, net, method="difference", null=NULL, threshold=0.05)
```



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```

+ attr: name (v/c), pagerank (v/n), pvalue (e/n)
+ edges from 396a8e9 (vertex names):
[1] PPM1G ->AATF   CTBP2 ->APP   TAGLN ->APP   MTSS1 ->APP   JMJD1C->ARID4A

```

3.3 Session Information

```
> sessionInfo()
```

```

R version 4.1.1 (2021-08-10)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Ubuntu 20.04.3 LTS

```

```

Matrix products: default
BLAS:   /home/biocbuild/bbs-3.14-bioc/R/lib/libRblas.so
LAPACK: /home/biocbuild/bbs-3.14-bioc/R/lib/libRlapack.so

```

```

locale:
 [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
 [3] LC_TIME=en_GB            LC_COLLATE=C
 [5] LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
 [7] LC_PAPER=en_US.UTF-8     LC_NAME=C
 [9] LC_ADDRESS=C             LC_TELEPHONE=C
[11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

```

```
attached base packages:
```

```
[1] stats4      stats      graphics  grDevices  utils      datasets  methods
[8] base
```

other attached packages:

```
[1] BSgenome.Hsapiens.UCSC.hg19_1.4.3
[2] BSgenome_1.62.0
[3] rtracklayer_1.54.0
[4] Biostrings_2.62.0
[5] XVector_0.34.0
[6] motifmatchr_1.16.0
[7] TFBSTools_1.32.0
[8] JASPAR2018_1.1.1
[9] annotate_1.72.0
[10] XML_3.99-0.8
[11] org.Hs.eg.db_3.14.0
[12] TxDb.Hsapiens.UCSC.hg19.knownGene_3.2.2
[13] GenomicFeatures_1.46.0
[14] AnnotationDbi_1.56.0
[15] GenomicRanges_1.46.0
[16] GenomeInfoDb_1.30.0
[17] IRanges_2.28.0
[18] S4Vectors_0.32.0
[19] bcellViper_1.29.0
[20] Biobase_2.54.0
[21] BiocGenerics_0.40.0
[22] pageRank_1.4.0
```

loaded via a namespace (and not attached):

```
[1] bitops_1.0-7          matrixStats_0.61.0
[3] DirichletMultinomial_1.36.0 bit64_4.0.5
[5] filelock_1.0.2       progress_1.2.2
[7] httr_1.4.2           tools_4.1.1
[9] utf8_1.2.2           R6_2.5.1
[11] seqLogo_1.60.0       DBI_1.1.1
[13] colorspace_2.0-2     prettyunits_1.1.1
[15] tidyselect_1.1.1     curl_4.3.2
[17] bit_4.0.4            compiler_4.1.1
[19] xml2_1.3.2           DelayedArray_0.20.0
[21] caTools_1.18.2      scales_1.1.1
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[83] restfulr_0.0.13	pracma_2.3.3
[85] tibble_3.1.5	GenomicAlignments_1.30.0
[87] memoise_2.0.0	ellipsis_0.3.2

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